STUDIES ON HELMINTH PARASITES FROM THE COAST OF FLORIDA. IV. DIGENETIC TREMATODES OF MARINE FISHES OF TAMPA, BOCA CIEGA BAYS, AND THE GULF OF MEXICO. 3 *

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Unless otherwise cited, all measurements are in millimeters.

Family Bucephalidae Poche, 1907

1. Rhipidocotyle transversale Chandler, 1935 (Figs. 1 to 2)

Host: Strongylura timucu (Walbaum) sp. inq.; needlefish; new host record?: family Belonidae Incidence of Infection: In 1 of 3 hosts

Number: One

Location: Mid-intestine

Locality: Bayboro Harbor, Tampa Bay, Florida; new lo-

cality record

Discussion: The identity of the host reported above is in question. There appear to be two common species of Strongulura in Bayboro Harbor. The hosts collected in this study were the finer scaled of the two forms and in addition possessed scaled cheeks. The identification is consistent with common usage as by Kilby (1955) and Longley & Hildebrand (1941). Hopkins (1954) reported R. transversale from Strongylura marina. Since the identity of S. marina, S. notata and S. timucu is somewhat confusing, we cannot be absolutely sure of Hopkin's report as well as our own. So far as the literature is concerned, this report of R. transversale

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is apparently the first from *Strongylura timucu* and also represents a new locality record. According to Manter (1947), Linton's (1940) *Prosorhynchus gracilescens* from Woods Hole is *R. transversale*, a species whose known distribution now includes Massachusetts, Virginia, Florida, Louisiana and Texas.

2. Bucephaloides bennetti Hopkins & Sparks, 1958 (Figs. 3 to 5)

Host: Paralichthys albiguttus Jordan & Gilbert; gulf fluke; new host record; family Pleuronectidae

Incidence of Infection: In 2 of 2 hosts

Numbers: 2, 42

Location: Pyloric ceca

Locality: Tarpon Key, Boca Ciega Bay, Florida; new locality record

Discussion: Manter (1954a.: p. 2) and Hopkins & Sparks (1958) indicated that Bucephalopsis bennetti Melugin, 1940, was a nomen nudum. The species was described in a thesis by Melugin and later given a name in abstract but no description included. Hopkins & Sparks (1958) retained the specific name bennetti for this species of Melugin. Hopkins (1954) named the genus Bucephaloides for all species previously in Bucephalopsis and retained the latter genus for B. haimeanus La Caze-Duthiers, a larval form. Bucephaloides bennetti was originally described from Paralichthys lethostigmus a species which appears to morphologically differ from Paralichthys albiguttus only in finray count. Hopkins and Sparks (1958) apparently did not describe the "lip" (fig. 5) of B: bennetti. Manter and Van Cleave (1951) described a closely related species, B. labiatus from a flounder, Paralichthys californicus, as possessing such a lip-like structure associated with the mouth. It is of some interest to note that the "lip" associated with the mouth has only been reported in Bucephaloides spp. from flounders of the genus Paralichthys even though sometimes widely separated geographically.

> 3. Bucephaloides caecorum Hopkins, 1956 (Figs. 6 to 7)

Host: Bairdiella chysurus (Lacepede); silver-perch; family Sciaenidae



Incidence of Infection: In 1 of 9 hosts

Number: One

Location: Pyloric cecum

Locality: Tarpon Key, Boca Ciega Bay, Florida; new lo-

cality record

Discussion: Bucephaloides caecorum has been reported from Cynoscion nebulosus and Bairdiella chrysurus by Hopkins (1956). Sparks (1958) records the parasite from the same hosts and Micropogon undulatus in Grand Isle, Louisiana. Bucephaloides caecorum is possibly an accidental parasite of B. chrysurus judging from the number of hosts found negative combined with the finding of only one worm. Bucephaloides caecorum is similar to B. bennetti, a form from which it may be instantly separated by the fact that the uterus of the latter does not extend so far anteriorly. the oral sucker is proportionately smaller, and an oral "lip" is present.

EXPLANATION OF FIGURES

Unless otherwise stated, all figures were drawn with the aid of a camera lucida. Scale has value indicated in millimeters. Abbreviations used: LC, Laurer's canal; OD, oviduct; OO, ootype; P, papilla or "lip"; USR, uterine seminal receptacle; UT, uterus.

Rhipidocotyle transversale Chandler, 1935, ventral view. Figure 1.

Figure 2. Same, eggs.

Figure 3. Bucephaloides bennetti Hopkins & Sparks, 1958, ventral view.

Figure

Same, mouth showing lip, pharynx dotted in. Figure 5.

6. Bucephaloides caecorum Hopkins, 1956, ventral view. Figure

7. Figure

Figure 8. Opecoeloides fimbriatus (Linton, 1934), ventro-lateral view.

Figure 9. Pseudopecoelus manteri (this paper), ventral view.
Figure 10A & B. Tormopsolus filiformis (this paper), scale drawing from camera lucida drawing of a twisted specimen, ventral view. Stephanostomum imparaspine (Linton, 1905) Manter, 1940, ven-Figure 11.

Same, ventral view of oral sucker and spines. Figure 12. Manteria brachyderus (Manter, 1940) Caballero, 1950, scale draw-Figure 13. ing from camera lucida drawing of a twisted specimen, ventral

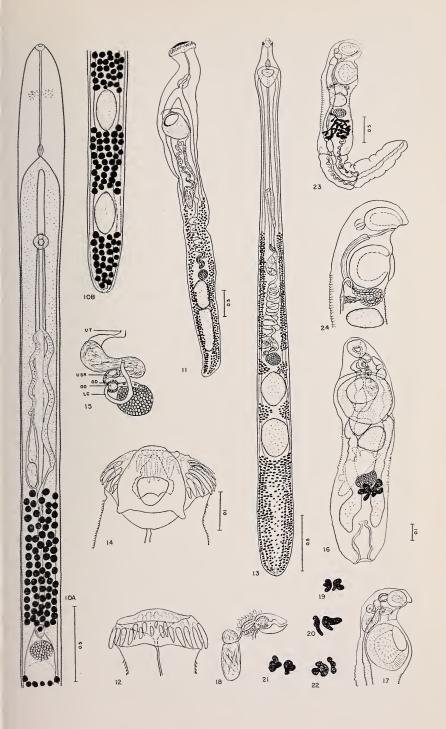
Same, ventral view of oral sucker and spines. Figure 14.

Same, mehlis complex.

Figure 15. Figure 16. Lecithochirium microstomum Chandler, 1935, ventral view.

Same, sketch of lateral view of anterior end of body. Figure 17.

Figure 18. Same, sketch of terminal genitalia.
Figure 19 to 22. Same showing variation in vitelline shape.
Figure 23. Dinurus tornatus (Rudolphi, 1899) Looss, 1907, lateral view.
Figure 24. Same, enlarged sketch of anterior end of body showing anterior extent of cuticular denticulations and posterior extent of hermaphroditic duct.



This report of B. caecorum from Florida represents a new locality record.

Family Opecoelidae Ozaki, 1925

4. Opecoeloides fimbriatus (Linton, 1934) Sogandares & Hutton (in press)
(Fig. 18)

Host: Bairdiella chrysurus (Lacepede); silver-perch; family Sciaenidae

Incidence of Infection: In 1 of 9 hosts

Number: One

Location: Pyloric cecum

Locality: Tarpon Key, Boca Ciega Bay, Florida

Discussion: Sogandares & Hutton (in press) have reported what they believe to be the metacercaria of O. fimbriatus encysted in the cephalothoracic cavity of the shrimp, Penaeus duorarum Burkenroad. Since this report, we have found a single adult of O. fimbriatus in a pyloric cecum of Bairdiella chrysurus.

The specimen of *O. fimbriatus* pictured in Figure 7 has a contracted anterior and posterior body. Aside from the fact that a uroproct is formed, the specimen is believed to be conspecific with the holotype of *O. fimbriatus* (USNM Helm. Coll. No. 8266) which we have examined. The acetabular papillae in the holotype of *O. fimbriatus* are somewhat retracted.

Linton (1905) pictures a specimen of *O. fimbriatus* (Fig. 178) from Beaufort, North Carolina. Hopkins (1941) reported both *O. fimbriatus* and *O. vitellosus* in *Bairdiella chrysurus* from the same locality. Sparks (1958) examined *Bairdiella chrysurus* in Grand Isle, Louisiana, but did not report *O. fimbriatus* from that area. Precluding the chance that *O. polynemi* Von Wicklen, 1946, is not a synonym of *O. fimbriatus*, this report appears to be the northernmost record of the species in the Gulf of Mexico.

5. Pseudopecoelus manteri n. sp. (Fig. 9)

Host: Bairdiella chrysurus (Lacepede); silver-perch; family Sciaenidae

Incidence of Infection: In 4 of 9 hosts

Number: 1, 1, 1, 1

Location: Pyloric ceca

Locality: Tarpon Key, Boca Ciega Bay, Florida Holotype: U. S. Nat. Mus. Helm. Coll. No. 39002

Diagnosis (Based on 4 specimens): Pseudopecoelus; body elongate, plump, broadly rounded posteriorly, conical anterior to acetabulum, 0.94 to 1.44 long by 0.42 to 0.56 wide. Cuticle unspined. Oral sucker terminal, 0.06 to 0.08 long by 0.10 to 0.12 wide. Acetabulum in anterior ½ body, short pedunculate, unornamented, 0.18 to 0.20 long by 0.16 to 0.22 wide. Sucker ratio 1:0.9 to 1.1. Prepharynx from ¼ length to approximately same length as pharynx. Pharynx 0.05 to 0.07 long by 0.06 to 0.07 wide. Esophagus approximately same length of pharynx. Cecal bifurcation immediately preacetabular; ceca ending blindly 3/3 distance between posterior testis and posterior end of body. Genital pore sinistral and level with anterior border of pharynx. Testes 2, intercecal, roundish, oblique, almost in contact with each other, anterior testis equatorial, sinistral to midline of body, 0.14 to 0.18 long by 0.16 to 0.18 wide; posterior testis dextral or median, 0.14 to 0.22 long by 0.12 to 0.20 Seminal vesicle from slightly posterior to acetabulum to enter a prostatic vesicle which is approximately same length of pharynx and connects with a very short cirrus which opens through genital pore. Ovary either with a single anterior notch or smooth, roundish to subtriangular in shape, to the right and level with anterior testis; 0.06 to 0.14 long by 0.10 to 0.12 wide. Seminal receptacle absent. Vitellaria medianly coalescing anterior and dorsal to acetabulum, extending posteriorly on each side of the body, overlapping ceca dorsally and ventrally to fill posttesticular space dorsally and ventrally. Uterus preovarian, extending from ovary to genital pore. Eggs collapsed, 48 to 60 microns long by 28 to 36 microns wide. Excretory pore terminal or subterminal, depending upon contraction of posterior end of body. Excretory vesicle tubular, extending from excretory pore to posterior level of ovary.

The name *manteri* is in honor of Professor Harold Winfred Manter, Department of Zoology, University of Nebraska, in recognition of his extensive contributions to the field of helminthology.

Discussion: This species is identified as P. umbrinae Manter & Van Cleave, 1951 and P. gibbonsiae Manter & Van Cleave, 1951 in Manter's (1954b: p. 6) key. Both species are from the coast of

California. Pseudopecoelus manteri differs from P. umbrinae by possessing smaller testes, an ovary which is never 3 to 4 lobed, and longer prostatic vesicle. Pseudopecoelus manteri differs from P. gibbonsiae by possessing a seminal vesicle which extends only slightly posterior to acetabulum, oblique testes, and genital pore at anterior border of pharynx, as compared with seminal vesicle extending far beyond acetabulum, almost to ovary, tandem testes and genital pore at mid-pharyngeal level. Pseudopecoelus manteri differs from all other species of Pseudopecoelus by possessing vitellaria which extend anterior to acetabulum as compared with vitellaria posterior to acetabulum.

Pseudopecoelus manteri in this collection was found associated with Bucephaloides caecorum Hopkins, 1956 and Tergestia pectinata (Linton, 1905) Manter, 1940.

Family Acanthocolpidae Lühe, 1906

6. Tormopsolus filiformis n. sp. (Figs. 10A, 10B)

Host: Rachycentron canadus (Linn.); cobia; family Rachycentronidae

Incidence of Infection: In 1 of 1 host

Number: 5

Location: Rectum

Locality: Gulf of Mexico, 8 miles offshore from John's

Pass, Madeira Beach, Florida

Holotype: U.S.N.M. Helm. Coll. No. 39003

Diagnosis (based on 5 specimens): Tormopsolus; body filiform, mainly cylindrical, 4.85 to 5.65 long by 0.20 to 0.36 wide at acetabulum. Cuticle spined to level of testes. Diffuse "eye-spots" present at mid-pharyngeal level. Forebody 0.90 to 1.30 long. Oral sucker terminal, 0.04 long by 0.05 to 0.06 wide. Acetabulum in anterior ¼ body, 0.09 by 0.09 to 0.10 wide. Prepharynx half length of forebody. Pharynx 0.07 to 0.08 long by 0.04 to 0.05 wide. Esophagus absent. Ceca extending from cecal bifurcation, one on each side of body to end at posterior end of body in a uroproct. Genital pore median, immediately pre-acetabular, followed by a long unspined muscular hermaphroditic duct which extends intercecally from genital pore, dorsal to acetabulum, to end approximately in anterior ½ body. Testes 2, intercecal, in tandem in posterior ¼

body; anterior testis 0.12 to 0.28 long by 0.16 to 0.18 wide; posterior testis 0.30 to 0.32 long by 0.16 wide. Cirrus sac mainly intercecal, dextral, extending from about equator of body to enter hermaphroditic duct; containing a heavily spined cirrus which is slightly longer than the bipartate internal seminal vesicle. Ovary intercecal, in posterior ½ body, 0.12 to 0.14 long by 0.12 to 0.14 wide. Vitellaria completely surrounding body ventrally and dorsally from posterior tip of cirrus sac, interrupted at level of ovary and each testis, to end at posterior end of body. Uterus extending intercecally from ovary to a heavily spined metraterm which is about same size of cirrus sac and connects with posterior portion of the hermaphroditic duct and cirrus sac junction. Receptaculum seminis uterinum present. Anterior extent of excretory vesicle not observed.

The name *filiformis* is from the latin *filum* (= hair)-*forma* (= form) for the hair-like appearance of the body of this species.

Discussion: Prior to our description of T. filiformis there were three species in the genus Tormopsolus Poche, 1926. These species are T. osculatus (Looss, 1901) Poche, 1926; T. orientalis Yamaguti, 1934 and T. lintoni Caballero, 1952. Tormopsolus filiformis differs from these species by possessing a hermaphroditic duct which extends far posterior to the acetabulum (almost ¼ distance from acetabulum to the anteriormost reach of the vitellaria) as compared with at most, hermaphroditic duct extending only a short distance posterior to acetabulum. Tormopsolus filiformis further differs from T. osculatus and T. orientalis by possessing a much longer forebody (proportionately twice as long) and longer prepharynx; and from T. lintoni by possessing an ovary which is separated from the anterior testis by a band of vitellaria.

7. Stephanostomum imparaspine (Linton, 1905) Manter, 1940 (Figs. 11 to 12)

Host: Rachycentron canadus (Linn.); cobia; family Rachycentronidae

Incidence of Infection: In 1 of 1 hosts

Numbers: 5

Location: Rectum

Locality: Gulf of Mexico, 8 miles offshore from John's Pass, Madeira Beach, Florida; new locality record.

Discussion: Linton (1905) described Distomum imparaspine from Rachycentron canadus in Beaufort, North Carolina. To our knowledge, the species has not since been reported in the literature until Manter (1940) placed D. imparaspine in the genus Stephanostomum.

Our specimens possessed from 33 to 34 oral spines. S. imparaspine resembles S. hispidum but differs in the number of crown spines (33 to 34 as compared with 40 to 42), and ovary always in contact with anterior testis as compared with ovary separated from anterior testis by a band of vitelline follicles.

It has been the current practice of ichthyologists to use the name Rachycentridae for the type genus *Rachycentron*. The name should be Rachycentronidae as here used.

8. Manteria brachyderus (Manter, 1940) Caballero, 1950 (Figs. 13 to 15)

Host: Oligoplites saurus (Bloch & Schneider); family Carangidae

Incidence of Infection: In 1 of 3 hosts

Numbers: 2

Location: Rectum

Locality: Bayboro Harbor, Tampa Bay, Florida; new lo-

cality record

Discussion: Manter (1940) originally placed this species in the genus Dihemistephanus. Caballero (1950) erected the genus Manteria for this species of Manter. Yamaguti (1953) placed the species in the genus Dihemistephanus and considered Caballero's (1950) Manteria, a sub-genus. Manteria brachyderus cannot be included in Dihemistephanus because D. lydiae, genotype, possesses a seminal receptacle. We have in our possession, 2 specimens of D. lydiae collected by Professor Manter from Mola mola in New Zealand and the seminal receptacle is definitely present. The presence of the seminal receptacle in D. lydiae would probably place it in the Deropristinae Cable & Hunnienen, 1942 of the Lepocreadiidae. The structure of the oral sucker, ventrally interrupted spines and an anterior acetabulum would place Manter's (1940) Dihemistephanus brachyderus in a genus all its own as proposed by Caballero (1950). For the present time, we prefer to retain Manteria Caballero, 1950. Our specimens possessed 30 and 32 oral spines respectively. Ather Siddiqui (personal communication), Purdue University, has studied specimens of *M. brachyderus* collected by Professor Raymond Cable, of the same institution, in Puerto Rico. He states that his immature specimens possess about 35, and mature specimens 41 to 47 oral spines. Professor Manter made a paratype of *M. brachyderus* available for our study. His specimen had 50 oral spines. We also have in our possession a specimen of *M. brachyderus* collected by Miss Margarita Bravo-Hollis, Instituto Nacional de Biologia (Mexico), in the Mexican Pacific. The oral spines have been lost, but the specimen is otherwise in good condition. Based on the material available for our study, we hesitate to name a new species for the Atlantic *Manteria* only on the basis of numbers of oral spines.

Caballero (1950) considers Linton's (1940) record of *Stephanosto-mum* sp. from Woods Hole, Massachusetts to be the same species as *M. brachyderus*. This record is the first from the Gulf of Mexico.

Family Hemiuridae Lühe, 1901

9. Lecithochirium microstomum Chandler, 1935 (Figs. 16 to 22)

Host: Coryphaena equisetis? Linn.; small dolphin; new host record; family Coryphaenidae

[ncidence of Infection: In 1 of 3 hosts

Numbers: 6

Location: Stomach

Locality: Gulf of Mexico, 15 mi. west of Pass-a-Grille,

Florida

Discussion: The specific identity of the host was not definitely established. The worms were dropped into cold A.F.A. fixative and the ecsoma of all specimens was retracted. Manter (1947) has pointed out that Chandler's (1935) figure of L. microstomum does not show a membrane surrounding the posterior end of the cirrus sac. Figure 18 is a sketch of the appearance of the terminal genitalia in our specimens. Figures 19 to 22 show observed variation of vitelline shape in our material.

Lecithochirium microstomum is known from the stomachs of many unrelated fishes in the American Atlantic and Tropical American Pacific. This report is the first from a fish of the genus Coryphaena.

10. Dinurus tornatus (Rudolphi, 1899) Looss, 1907 (Figs. 23 to 24)

Host: Coryphaena equisetis? Linn.; small dolphin; family Corvphaenidae

Incidence of Infection: In 3 of 3 hosts

Numbers: Not counted Location: Stomach

Locality: Gulf of Mexico, 15 miles west of Pass-a-Grille,

Florida

Discussion: The specific identity of the host is in question. All three hosts were collected from a single school of small dolphins.

Manter (1947) reported D. tornatus from Coryphaena hippurus in Tortugas, Florida, and Linton (1905) from Coryphaena hippurus and C. equisetis in Beaufort, North Carolina. Ward (1954) reported "D. tornatus" from the Miami, Florida area. She followed Dawes (1947) in considering D. longisinus Looss, 1907, a synonym of D. The species Ward (1954) pictures (Fig. 3) may be D. longisinus because the hermaphroditic duct extends to the middle of the acetabulum. Dawes (1947) considers Dinurus barbatus (Cohn, 1903) Looss, 1907, D. breviductus Looss, 1907, D. longisinus Looss, 1907, synonyms of D. tornatus (Rudolphi, 1899) Looss, 1907. One of us (F.S.) has Dinurus spp. collected from Coryphaena hippurus in the Gulf of Panama and in Bimini, B.W.I., and in addition has been privileged to study Professor Manter's series of Dinurus spp. from Coryphaena hippurus collected in Tortugas, Florida and the Tropical American Pacific. Dinurus tornatus may be easily separated from D. longisinus because the cuticle in the former species is denticulated to the acetabulum (Figs. 23 to 24) and the hermaphroditic duct extends almost to or beyond the posterior border of the acetabulum, as compared with cuticular denticulations extending to the oral sucker and hermaphroditic duct extending to about the middle of the acetabulum in D. longisinus. These differences appear to be constant in several hundred specimens which one of us (F.S.) has been able to observe. The fact that most Dinurus spp. may live together in a single host is not evidence enough for synonymy of the species as presented by Dawes (1947). There are numerous examples to indicate that closely related species and genera may cohabit in the same host. The finding of closely related species living together in one host would imply a strong degree of host specificity accompanying evolution. Manter (1957) shows how the accacoelid trematodes of Mola mola are a good example of such evolution. There is no definite contrary evidence to show that hermaphroditism is the rule in the Digenea. There is some evidence that cross fertilization occurs, and it may be more common than we at present suspect or assume. The differences of the cuticular denticulations of Dinurus tornatus and D. longisinus could possibly be a recognition character preventing interspecific mating if cross fertilization occurs in this group of trematodes. Thus in trematodes in which weak recognition characters preventing interspecific matings would exist, hybrids would be expected to result. Pseudocreadium myohelicatum (Bravo & Manter, 1957) seems to be just such a hybrid example (Pseudocreadium scaphosomum Manter, 1940 X Pseudocreadium spinosum Manter, 1940). Such a hybridization may perhaps occur when two trematode species normally occur in separate host species and sometimes are accidentally in one host species together. Recognition characters preventing interspecific matings in these instances would possibly be weaker than when both species normally occur together in one host species as is the case in the Dinurus spp. in Coryphaena spp., in the monorchiids from Haemulon spp., and in the accacoelids from Mola mola. Price (1953) believed hybridization between Faciola hepatica and F. gigantica existed in cattle in the Southeastern United States. If trematode hybrids exist these would probably be confused with intergradation between two closely related species by most observers. Such may be the case in some *Dinurus* spp. confused as synonyms by some investigators.

Summary

- 1. Two new trematodes are named and described. These species are: *Pseudopecoelus manteri* (family Opecoelidae) from *Bairdi- 'ella chrysurus* in Boca Ciega Bay, Florida, and *Tormopsolus fili- formis* (family Acanthocolpidae) from *Rachycentron canadus* in the Gulf of Mexico.
- 2. New host records are as follows: Rhipidocotyle transversale Chandler, 1935 (family Bucephalidae) in Strongylura timucu?; Bucephaloides bennetti Hopkins & Sparks, 1958 (family Bucephalidae) in Paralichthys albiguttus; and Lecithochirium microstomum Chandler, 1935 in Coryphaena equisetis?.

- 3. New locality records are: From Tampa and/or Boca Ciega Bays, Rhipidocotyle transversale Chandler 1935; Bucephaloides bennetti Hopkins & Sparks, 1958; and Bucephaloides caecorum Hopkins, 1956 (family Bucephalidae) Manteria brachyderus (Manter, 1940) Caballero, 1950 (family Acanthocolpidae); from the Gulf of Mexico, Stephanostomum imparaspine (Linton, 1905) Manter, 1940 (family Acanthocolpidae).
- 4. Information pertaining to the exact location and numbers of each trematode species within its respective host(s) is given when possible.

REFERENCES CITED

CABALLERO yC., E.

1950. Un nuevo genero de trematodo de peces marinos perteneciente a la familia Acanthocolpidae, 1909. Ann. Inst. Biol. (Mexico), 21(1): 95-102.

CHANDLER, A. C.

1935. Parasites of fishes in Galveston Bay. Proc. U. S. Nat. Mus., 83: 123-157.

DAWES, B.

1947. The trematoda of British Fishes. The Ray Society, 364 pp.

KILBY, J. D.

1955. The fishes of two gulf coastal marsh areas of Florida. Tulane Stud. Zool., 2(8): 175-247.

LONGLEY, W. H., and S. F. HILDEBRAND

1941. Systematic catalogue of fishes of Tortugas, Florida. Carnegie Inst. Wash. (Pub. 535), Papers Tortugas Lab., 34: 1-331, 34 pls.

HOPKINS, S. H.

- 1941. Excretory systems of Helicometra and Cymbephallus (Trematoda), with remarks on their relationships. Trans. Am. Micr. Soc., 50(1): 41-44.
- 1954. The American species of trematode confused with *Bucephalus (Bucephalopsis)haeimeanus*. Parasitol., 44(3&4): 353-370.
- 1956. Two new trematodes from Louisiana, and the excretory system of Bucephalidae. Trans. Am. Micr. Soc., 75(1): 129-135.

HOPKINS, S. H., and A. K. SPARKS

1958. A new species of *Bucephaloides* (Trematoda; Bucephalidae) from a marine fish of Grand Isle, Louisiana. J. Parasitol., 44(4): 409-411.

LINTON, E.

1905. Parasites of fishes of Beaufort, North Carolina. Bull. Bur. Fish. 24: 321-428, 34 pls.

1940. Trematodes from fishes mainly from the Woods Hole region Massachusetts. Proc. U. S. Nat. Mus., 88: 11-72, 26 pls.

MANTER, H. W.

- 1940. Digenetic trematodes of fishes from the Galapagos Islands and neighboring Pacific. Allan Hancock Pac. Exp. 2(14): 329-497.
- 1947. Digenetic trematodes of marine fishes of Tortugas, Florida. Amer. Mid. Nat., 38(2): 257-416.
- 1954a. Trematoda of the Gulf of Mexico, in Gulf of Mexico its origin, waters, and marine life. Fish. Bull. U. S. Fish & Wildlife Service, 89: 335-350.
- 1954b. Some digenetic trematodes from fishes of New Zealand. Trans. Roy. Soc. N. Z., 82(2): 475-568.
- 1957. Host specificity and other host relationships among the digenetic trematodes of marine fishes. First Symposium on host specificity among parasites of vertebrates (Neuchatel): 185-196.

MANTER, H. W., and H. VAN CLEAVE

1951. Some digenetic trematodes, including eight new species, from marine fishes of La Jolla, Calif., Proc. U. S. Nat. Mus., 101: 315-340, 2 pls.

PRICE, E.

1953. The fluke situation in American ruminants. J. Parasitol., 39: 119-134.

SOGANDARES-BERNAL, F., and R. F. HUTTON

(In Press) Studies on helminth parasites from the coast of Florida. III. Digenetic trematodes of marine fishes from Tampa and Boca Ciega Bays. 2. J. Parasitol.

SPARKS, A. K.

1958. Some digenetic trematodes of fishes of Grand Isle, Louisiana. Proc. Louisiana Acad. Sci., 20: 71-82.

WARD, H.

1954. Parasites of marine fishes of the Miami region. Bull. Mar. Sci. Gulf and Carib., 4(3): 244-261.

YAMAGUTI, S.

1953. Systema Helminthum. Part I. Digentic trematodes of fishes. Maruzen Co. Ltd., Nihobashi, Tokyo, Jayan.